Fortessa X20

# Flow Cytometry & Single Cell Analysis https://fcsc.ku.dk/

												Secondary I	lourochr	ome (Bright	t Antigen)													
	BUV395	BUV496	BUV737	BV421	Pac Blue	BV480	BV510	BV605	BV650	BV711	BV786	FITC	AF 488	BB515	PerCP-Cy5.5	BB700	PE	PE-C	CF594 PE	-Cy5 I	PE-Cy7	APC	AF 647	AF 700	APC-R700	APC-Cy7	APC-Fire	APC-eFlour780
BUV395	154	48%	479	<mark>6</mark> 5%	5 9	% 119	6 89	6%	8%	% 79	6 85	% <mark>7</mark> 9	5 55	69	% 9	% 8%	6	8%	6%	14%	6%	4%	7%	8%	79			8%
BUV496	68%	157	339	6 <mark>49%</mark>	22	% 83%	6 769	6 18%	16%	% 149	6 169	<mark>%</mark> 23%	5 15	6 39%	% 21	% 16%	61	16%	17%	29%	19%	20%	12%	19%	13%	6 26%	6 17%	23%
BUV737	6%	27%	33	1 4%	5 2	.% 16%	6 349	68%	66%	6 719	669 669	<mark>%</mark> 6%	5 49	% 49	<mark>%</mark> 62	% 73%	62	23%	<mark>41%</mark>	75%	47%	59%	17%	35%	68%	6 38%	6 30%	30%
BV421	16%	16%	229	6 510	)	199	6 209	6 <mark>46%</mark>	48%	<mark>% 45</mark> %	<mark>67</mark> 5	<mark>%</mark> 18%	5 179	6 22%	% 26			21%	26%	63%	20%	20%	20%	24%	25%	6 29%	6 26%	
Pacific Blue	15%				-	12 19%	6 19%		47%					_	% 26			21%	27%	63%	20%	20%		24%	25%			
BV480	31%							29%										32%	32%	43%	33%	32%		42%	30%			
E BV510	31%		-				4											32%	32%	43%	33%	32%	_	42%	30%			
.eg BV605	11%					% <mark>60</mark> %	-	6 164										5 <mark>9%</mark>	66%	73%	18%	33%		22%	49			and the second se
BV650	5%	18%				% <mark>47</mark> %		<mark>6</mark> 85%	79			_						5 <mark>3%</mark>	68%	93%	21%	71%		11%	229	-		
E BV711	5%	14%				<b>1%</b> 40%		<u> </u>		6 4		_						52%	69%	91%	24%	60%		25%	54%			
BV786	3%					<mark>%</mark> 33%		<mark>6 79%</mark>			6 30			% 5%			-	27%	45%	75%	80%	51%		33%	60%			
E FITC	21%														27			30%	24%	39%	25%	26%	23%	32%	29%			
Alexa Fluor 488	21%												5		27			81%	24%	39%	25%	26%		32%	299			
BB515	21%							6 20%						15				31%	24%	39%	25%	26%		32%	299			
PerCP-Cy5.5	5%			6 2%		1% 9%		6 5/%	41%							80		71%	84%	96%	46%	66%		21%	389			
BB700	5%			6 2%	_	% 9%			41%		-					201		/1%	84% 84%	97%	47%	66%	8%	21%	389		-	•
E PE PE-CF594	18% 10%	18%							29%	% 249 <mark>%</mark> 129								553 2%	463	94%	54% 37%	20% 22%		23% 13%	239			
PE-Cr594 PE-Cy5		13%		-					44%	1								72%	85%	66% 336	37% 14%	77%			159 259			
PE-Cy5 PE-Cy7	1% 2%	1%				%		62%		6 <u>12</u> 9 6349								51%	85%	330	424	70%		13% 45%	789			
APC	2%	1%				% 3/ % 12%		6 38%									_	85%	58%	97%	12%	169		24%	749			
Alexa Fluor 647	0%	1%				/% 12/ 1% 129		6 38%										85%	58%	97%	12%	105	81	24%	749			
Alexa Fluor 700	0%	1%		6 0%		% 10%			61%									9%	24%	90%	26%	88%		24/0	-	58%		
APC-R700	0%	1%	879	6 0%		% <u>10</u> %			61%									9%	24%	90%	26%		82%	22	18			
APC-Cv7	0%	3%		6 0%		% 107 % 49		6 5%	34%									2%	6%	79%	70%	77%	68%	58%	839		9	50/8
APC-Fire	0%	3%		6 0%		.% 4%		6 5%	34%									2%	6%	79%	70%	77%		58%	839	6	72	
APC-eFlour780	0%			6 0%		.% 49			34%									2%	6%	79%	70%	77%		58%	839	6	,,	74

#### How the matrix was made

Mouse spleens was stained with individual anti-CD8 labeled antibodies with the indicated flourochrome and analyzed on the indicated instrument.

Each calculated value was arbitrarily assigned a color code according to the legend to show where the biggest spreading was situated.

#### How to use the resolution impact matrix

You find the color of interest on the top of the matrix, go down till you find the channel that you need to combine the color with and read the impact of spreading.

### Consider this for panel design

When you are designing larger panels the task of making correct combinations becomes more difficult, but using the list below can help you:

- The lineage markers such as CD4, CD19 etc. should be found on the top of the matrix.
- Make sure the lineage marker has as many green cells as possible.
- For an important marker you should find the color on the left of the matrix.
- Make sure the marker has as many green cells as possible when you move across the matrix.
- Notice that spreading occurs between different laser lines.

## Amount of spreading (%)

0-20
20-40
40-60
60-80
80-100
NA

Relative fluorochrome brightness (AU)

